

EPPO Datasheet: *Potyvirus plumpxi*

Last updated: 2020-02-05

IDENTITY

Preferred name: *Potyvirus plumpxi*

Taxonomic position: Viruses and viroids: Riboviria: Orthornavirae: Pisuviricota: Stelpaviricetes: Patatavirales: Potyviridae

Other scientific names: *PPV*, *Plum pox potyvirus*, *Plum pox virus*, *Prunus virus 7*

Common names: pox of plum, sharka

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EPPO Categorization: A2 list

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EU Categorization: RNQP (Annex IV)

EPPO Code: PPV000



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Notes on taxonomy and nomenclature

PPV is so far the only potyvirus known to infect temperate fruit trees. The potential existence of a serologically related virus in some *Prunus* materials of Asian origin has been reported (Hadidi & Levy, 1994). The existence and identity of this virus, tentatively named prunus latent potyvirus has however not been confirmed in further efforts. In particular, High-Throughput Sequencing of several *Prunus* sources initially reported to be infected by the prunus latent potyvirus or showing similar PPV-cross reactions to it failed to identify any potyvirus or PPV-like virus (Marais *et al.*, 2016).

HOSTS

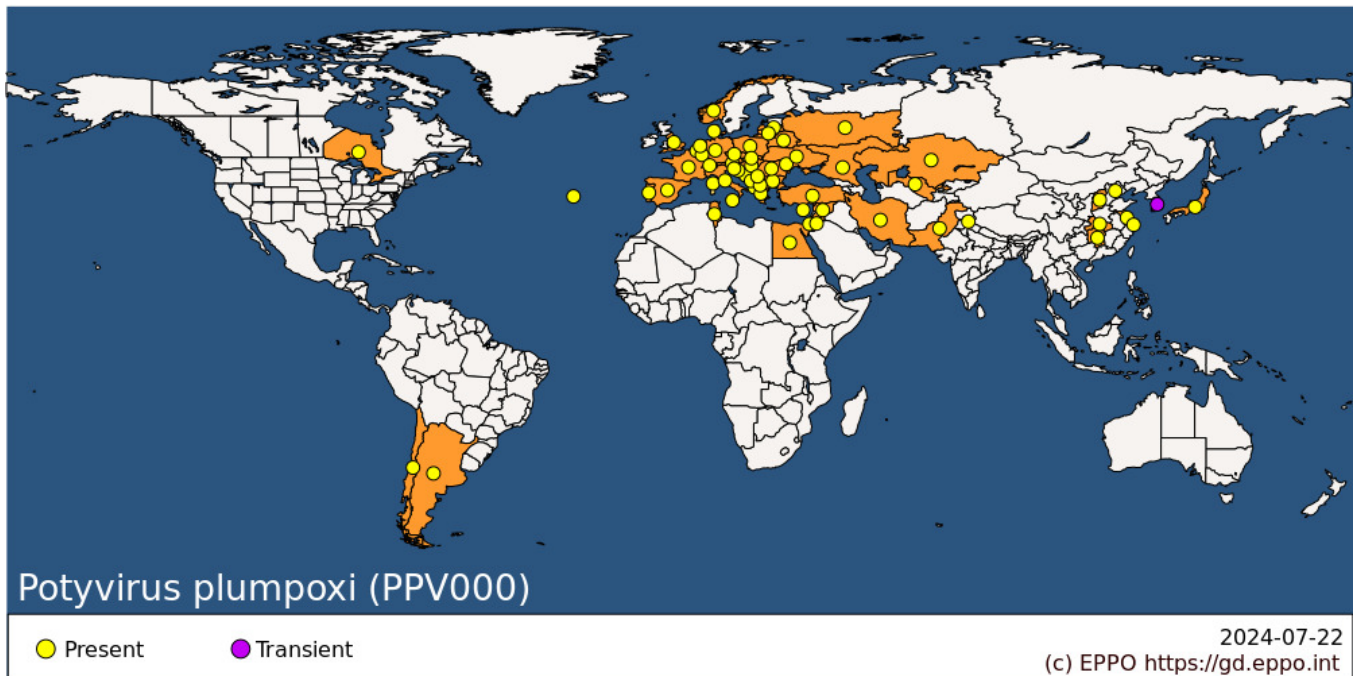
The main woody hosts are the species of *Prunus* grown for fruit production, including apricot (*P. armeniaca*), peach (*P. persica*) and plum (*P. domestica* and *P. salicina*). Almond trees (*P. dulcis*) can be infected by PPV but show few symptoms (Dallot *et al.*, 1997, Damsteegt *et al.*, 2007). Natural infection of *P. cerasus* and *P. avium*, attributed to the cherry adapted PPV-C strain has been sporadically observed in Europe (Kalashyan *et al.* 1994; Crescenzi *et al.*, 1997). The recent identification of two other cherry-adapted strains (PPV-CR and CV, Glasa *et al.*, 2013; Chirkov *et al.*, 2018) also shows the epidemiological potential of these PPV strains in the cherry hosts.

Many *Prunus* species used as rootstock or as ornamentals are natural hosts of PPV, together with a range of wild *Prunus* species, including their interspecific hybrids (James & Thompson, 2006; Damsteegt *et al.*, 2007). PPV infects most wild or ornamental species of *Prunus*, such as *P. besseyi*, *P. cerasifera*, *P. insititia*, *P. spinosa*, *P. tomentosa*, serving as a potential reservoir and source of virus inoculum. Numerous annual cultivated plants or weeds have been shown to be experimental hosts of PPV (Virsecq Marn *et al.*, 2004; Llacer, 2006). However, as reports of natural infection of such herbaceous hosts have never been confirmed using two independent diagnostic techniques, and sequence information on the isolate(s) involved has never been provided, their host status is unconfirmed. In any case, natural transmission between such herbaceous plants and *Prunus* has never been demonstrated in nature, so that the epidemiological contribution of herbaceous hosts, if any, remains questionable.

Host list: *Prunus americana*, *Prunus armeniaca*, *Prunus avium*, *Prunus besseyi*, *Prunus brigantina*, *Prunus cerasifera*, *Prunus cerasus*, *Prunus curdica*, *Prunus domestica* subsp. *insititia*, *Prunus domestica* subsp. *italica*, *Prunus domestica*, *Prunus dulcis*, *Prunus glandulosa*, *Prunus holosericea*, *Prunus incisa*, *Prunus japonica*, *Prunus laurocerasus*, *Prunus mahaleb*, *Prunus mandshurica*, *Prunus maritima*, *Prunus mume*, *Prunus nigra*, *Prunus persica*, *Prunus pumila*, *Prunus salicina*, *Prunus serotina*, *Prunus serrulata*, *Prunus sibirica*, *Prunus simonii*, *Prunus spinosa*, *Prunus tomentosa*, *Prunus triloba*, *Prunus virginiana*, *Prunus x blireana*, *Prunus x cistena*, *Prunus*, *Spiraea* sp., *Tilia*

GEOGRAPHICAL DISTRIBUTION

Typical sharka symptoms, caused by PPV (Atanasoff, 1932) were observed for the first time in plums in Eastern Europe (Bulgaria) around 1914. PPV subsequently spread, over most of the European continent and Mediterranean basin during the 20th century (Garcia & Cambra, 2007). PPV has also been reported from the Americas (Levy *et al.*, 2000; Thompson *et al.*, 2001; Herrera, 2013), from Asia (Maejima *et al.*, 2010) and from Africa (Boulila *et al.*, 2004). It is not yet officially reported from Oceania. In 2019, PPV was reported to be eradicated in the USA (USDA, 2019).



EPPO Region: Albania, Austria, Belarus, Belgium, Bosnia and Herzegovina, Bulgaria, Croatia, Cyprus, Czech Republic, Denmark, France (mainland, Corse), Germany, Greece (mainland), Hungary, Israel, Italy (mainland, Sicilia), Jordan, Kazakhstan, Latvia, Lithuania, Luxembourg, Moldova, Montenegro, Netherlands, North Macedonia, Norway, Poland, Portugal (mainland, Azores), Romania, Russia (Central Russia, Southern Russia), Serbia, Slovakia, Slovenia, Spain (mainland), Switzerland, Tunisia, Türkiye, Ukraine, United Kingdom (England), Uzbekistan

Africa: Egypt, Tunisia

Asia: China (Beijing, Hubei, Hunan, Jiangsu, Shanghai, Shanxi), India (Himachal Pradesh), Iran, Israel, Japan (Honshu), Jordan, Kazakhstan, Korea, Republic, Pakistan, Syria, Uzbekistan

North America: Canada (Ontario)

South America: Argentina, Chile

BIOLOGY

Infected *Prunus* trees are the major source of inoculum. The virus is transmitted from them either by grafting and other vegetative multiplication techniques or non-persistently by aphid vectors (Ng & Falk, 2006; Moreno *et al.*, 2009). *Aphis spiraecola*, *Phorodon humuli*, *Hyalopterus pruni* and *Myzus persicae* are the main vectors (Cambra & Vidal, 2017). Other aphids have also been shown to transmit the virus: *Aphis craccivora*, *A. fabae*, *A. gossypii*, *A. hederarum*, *Brachycaudus cardui*, *B. helychrysi*, *B. persicae*, *Myzus cerasi*, *M. varians*, *Rhopalosiphum padi* and *Sitobion fragariae* (Labonne *et al.*, 1995; Gildow *et al.*, 2004).

The number of trees becoming infected in an orchard is directly related, in a given season, to the population level of winged aphids. These aphids probe or feed on infected leaves, then fly to other trees where they again probe or feed (Labonne & Quiot, 2006). Aphids can also acquire PPV from infected fruits (Labonne & Quiot, 2001). Analysing the spatial distribution of aphid-borne spread in eastern Spain, Gottwald *et al.* (1995) concluded that aphids do not spread the disease much to immediately adjacent trees, but to a few trees away. Experiments and modeling show that spread occurs generally within a few hundred meters with about 50% of transmission events occurring within 90 m of the source tree (Pleydell *et al.*, 2018). The capacity for vector transmission can vary between viral isolates even

within the same strain (Dallot *et al.*, 2003; Glasa *et al.*, 2004). After inoculation of a *Prunus* tree, the incubation period may last several months and systemic spread may take several years. Accordingly, the virus may be distributed very irregularly in trees, possibly explaining the dynamic structure and heterogeneous nature of PPV population(s) in individual hosts (Jridi *et al.*, 2006; Predajza *et al.*, 2012). Seed or pollen transmission of PPV in *Prunus* has not been confirmed, and is unknown in practice (Glasa *et al.*, 1999; Pasquini & Barba, 2006).

Various strains of PPV were originally distinguished (necrotic, intermediate, yellow) on the basis of symptoms obtained by inoculation of herbaceous indicator plants (Sutic *et al.*, 1961). Then two isolates D (Dideron) and M (Markus), the former on apricot in France and the latter originally on peach in Greece, were serologically differentiated (Kerlan & Dunez, 1979). Further efforts led to the identification of these isolates as typifying two strains differing in serological and molecular properties (Candresse *et al.*, 1998). Later sequencing efforts led to the recognition of further strains (Wetzel *et al.*, 1991; Nemchinov *et al.*, 1996; Glasa *et al.*, 2004, Ulubas Serçe *et al.*, 2009; James & Varga A, 2005; Palmisano *et al.*, 2012; Glasa *et al.*, 2013, Chirkov *et al.*, 2018). Currently, a total of ten genetic strains are recognized for PPV (in the order of their discovery: D, M, EA, C, Rec, T, W, An, CR and CV). The three main strains, that have very wide geographical distributions, are PPV-M, D and Rec (Garcia *et al.*, 2014). Some strains have particular biological/epidemiological features (e.g. cherry-adapted strains C, CR and CV) or a restricted geographical distribution (EA in Egypt, T in Turkey). However, due to a high intra-strain variability, most of strains do not show clear-cut epidemiological characteristics that would separate them from others (Sihelská *et al.*, 2017). Several strains, including Rec and T have been shown to result from recombination events involving the D and M strains (Glasa *et al.*, 2004; Glasa & Candresse, 2005; Hajizadeh *et al.*, 2019).

DETECTION AND IDENTIFICATION

Symptoms

Symptoms may appear on leaves or fruits as a consequence of physiological, biochemical, proteomic, and transcriptional or post-transcriptional changes induced by viral infection (Clemente-Moreno *et al.*, 2015). The symptoms are particularly clear on leaves at the beginning of the vegetation period: chlorotic spots, bands or rings, vein clearing, or even leaf deformation in peaches. Infected fruits show chlorotic spots or rings. Diseased plums and apricots may be deformed and show internal browning of the flesh; in apricot, the stones show characteristic pale rings or spots. Premature fruit dropping (up to 100%) can occur in the most susceptible cultivars (Sochor *et al.*, 2012; Garcia *et al.*, 2014). Symptoms of sharka depend very much on PPV isolate, locality, season, *Prunus* species and cultivar and plant organ (leaf or fruit) (Dosba *et al.*, 1986).

Morphology

PPV has filamentous virus particles 750 nm long and 15 nm in diameter. It has a single-stranded RNA genome of ca 10 000 nucleotides, coding for a large polyprotein with a molecular weight of 3.5×10^6 Da. The genome encodes 10 mature proteins processed from the viral polyprotein by the action of three viral proteases. As for other potyviruses, transcriptional slippage allows the extension of an out of frame short open reading frame P3N-PIPO (Rodamilans *et al.*, 2015).

Protein inclusions of the pinwheel type are present in the cytoplasm of infected cells. The full-length nucleotide sequences of a number of virus isolates belonging to all recognized strains have been determined (García *et al.*, 2014). Genome function in PPV is now increasingly understood, and this virus is now a model for studies on the molecular biology of potyviruses (García *et al.*, 2014; Rodamilans *et al.*, 2019).

Detection and inspection methods

In spite of the irregular distribution of the virus in the tree, visual inspection may allow detection of symptoms in susceptible cultivars, especially during the period of active growth. Testing on susceptible indicators (peach GF305 or *Prunus tomentosa*) by chip-budding can produce symptoms in 6-8 weeks (Damsteegt *et al.*; 1997, Gentit, 2006). Mechanical inoculation on *Chenopodium foetidum* or *Nicotiana benthamiana* produces symptoms in 6-10 days but the inoculation efficiency from *Prunus* hosts is generally low (Sutic *et al.*, 1961; Glasa & Candresse, 2005; Glasa *et al.*, 2010).

Immunochemical methods, such as ELISA, have still an important role in the diagnostic of PPV (Šubr & Glasa, 2008; Cambra *et al.*, 2011). A range of broad-spectrum or strain-specific antibodies are available (Cambra *et al.*, 1994; Cambra *et al.*, 2006a; Candresse *et al.*, 2011), including monoclonal antibodies. Although all parts of the tree can be sampled for testing, the best detection results rely on the use of composite leaf samples from actively growing shoots taken in different parts of the canopy (Adams, 2008).

Molecular methods based on the amplification of specific parts of the PPV genome show a higher sensitivity than immunochemical methods (Lopez *et al.*, 2003). Various modifications of RT-PCR in single or multiplex format have been developed both for the universal detection of all PPV isolates or for strain-specific detection (Olmos *et al.*, 2002; Šubr *et al.*, 2004).

An effective detection coupled with the possibility to differentiate PPV strains can be achieved using real-time RT-PCR (Varga & James, 2005; Capote *et al.*, 2009; Fotiou *et al.*, 2019). Isothermal amplification methods, such as LAMP (Varga & James, 2006; Hadersdorfer *et al.*, 2011) have also been developed for a simple and direct use in the field. Validated international protocols for detection and characterization of PPV are available (EPPO, 2004, IPPC-FAO, 2012).

PATHWAYS FOR MOVEMENT

The distribution of the disease appears to be at random in orchards. The virus is introduced as a consequence of aphid transmission or of the use of infected planting material. After 2-3 years, infection begins to spread from the first infected trees. Graft transmission can contribute significantly to spread in infected areas if certified virus-free material is not used. Movement of the virus between areas or countries is most often linked to the use of uncertified plants for planting (Rimbaud *et al.*, 2015a, b).

PEST SIGNIFICANCE

Economic impact

The importance of sharka disease on the European stone-fruit production has been reviewed by Cambra *et al.* (2006b). The disease incidence is particularly high in the fruit-producing areas of central and eastern Europe. Virus infection can lead to considerable yield losses, reaching 100%. European plums may show premature fruit drop, while Japanese plums and peaches show ring-spotting on fruit, and apricots show serious fruit deformation.

Control

There is no anti-virus treatment available to control sharka disease in orchards. There are, however, considerable differences in susceptibility between the cultivars available for use in countries where infection is widespread (Kegler *et al.*, 1998, Martínez-Gómez *et al.*, 2000). However, the frequent plantation of tolerant *Prunus* cultivars (their fruits remaining generally symptomless in case of infection) has probably contributed to the further spread of PPV in these countries (Glasa *et al.*, 2004). Biological control by inoculation of trees with hypo-aggressive strains has not proved as successful in the field as under controlled conditions (Kerlan *et al.*, 1980) and is not considered a realistic preventative option. Other effective control methods are the production and use of healthy plants for planting within a certification system, and the eradication of diseased trees or orchards to reduce inoculum pressure (Rimbaud *et al.*, 2015a). As for other potyviruses, the control of aphid vectors by regular treatment with aphicides or mineral oils shows only limited effectiveness, with the possible exception of nurseries where some protection has been recorded (Vidal *et al.*, 2013). Such methods are used to contain PPV in several countries (e.g. France, Italy). EPPO recommends a certification scheme for fruit trees, which takes into account PPV (EPPO, 1991/1992). Resistance to PPV shows some promise, whether by traditional breeding or by transgenic methods. The hypersensitive response in plums, resulting in localized cell death, has been found to be an effective resistance mechanism against PPV (Hartmann, 1998). Apricot varieties resistant to the PPV-D strain are now extensively planted in some areas of Spain. While progress has been obtained in plum and apricot, the development of resistant peach varieties has remained a challenge due to the paucity of resistance sources. Biotechnology has also contributed

with the development of the transgenic plum cultivar Honeysweet which shows a high, broad spectrum resistance (Scorza *et al.*, 2016).

Phytosanitary risk

PPV is included in the EPPO A2 list of pests recommended for regulation as quarantine pests. It is a quarantine pest for the European Union and many other EPPO member countries. It is also of regulatory interest to other Regional Plant Protection Organizations (e.g. COSAVE, IAPSC and NAPPO).

In the EPPO region, PPV presents a major risk to apricot, plum and peach in many countries where it is still absent or very localized. In addition, its presence in a country creates difficulties for export of certified planting material.

PHYTOSANITARY MEASURES

In order to prevent entry or spread of PPV, all imported host material (except seeds) should come from a place of production subject to growing-season inspection (EPPO, 2016). If the virus is present in the exporting country, this inspection should also concern the immediate vicinity of the place of production, and the material should derive from tested mother plants. Material produced following the EPPO certification scheme for virus-free fruit trees would satisfy these requirements (EPPO, 1991/1992).

Measures can effectively be taken to prevent spread of PPV from foci of infection and even to eradicate it. These include planting non-host plants in infected areas, using tolerant or resistant cultivars, controlling the vectors and destroying all diseased trees.

REFERENCES

- Adams A (2008) The detection of plum pox virus in *Prunus* species by enzyme-linked immunosorbent assay (ELISA). *Annals of Applied Biology* **90**, 215-221.
- Atanasoff D (1932) Plum pox. A new virus disease. *Annals of the University of Sofia, Faculty of Agriculture and Silviculture* **11**, 49-69.
- Boulila M, Briard P, Ravelonandro M (2004) Outbreak of *Plum pox virus* in Tunisia. *Journal of Plant Pathology* **86**, 197-201.
- Cambra M, Vidal E (2017) Sharka, a vector-borne disease caused by *Plum pox virus*: vector species, transmission mechanism, epidemiology and mitigation strategies to reduce its natural spread. *Acta Horticulturae* **1163**, 57-68.
- Cambra M, Boscia D, Gil M, Bertolini E, Olmos A (2011) Immunology and immunological assays applied to the detection, diagnosis and control of fruit tree viruses. In: *Virus and Virus-like Disease of Pome and Stone Fruits* (Hadidi, A., Barba, M., Candresse, T. and Jelkmann, W., eds), pp. 303–313. St. Paul, Minnesota: APS Press.
- Cambra M, Boscia D, Myrta A, Palkovics L, Navrátil M, Barba M, Gorrís MT, Capote N (2006a) Serological detection and characterisation of *Plum pox virus*. *Bulletin OEPP/EPPO Bulletin* **36**, 254-261.
- Cambra M, Capote N, Myrta A, Llácer G (2006b) *Plum pox virus* and the estimated costs associated with sharka disease. *Bulletin OEPP/EPPO Bulletin* **36**, 202-204.
- Cambra M, Asensio M, Gorrís, M, Pérez E, Camarassa E, García JA, Moya JJ, López-Abella D, Vela C, Sanz A (1994) Detection of *Plum pox potyvirus* using monoclonal antibodies to structural and non-structural proteins. *Bulletin OEPP/EPPO Bulletin* **24**, 569-577.
- Candresse T, Cambra M, Dallot S, Lanneau M, Asensio M, Gorrís MT, Revers F, Macquaire G, Olmos A, Boscia D, Quiot JB, Dunez J (1998) Comparison of monoclonal antibodies and polymerase chain reaction assays for the typing of isolates belonging to the D and M serotypes of *Plum pox potyvirus*. *Phytopathology* **88**, 198-204.

- Candresse T, Saenz P, García JA, Boscía D, Navratil M, Gorris MT, Cambra M (2011) Analysis of the epitope structure of *Plum pox virus* coat protein. *Phytopathology* **101**, 611–619.
- Capote N, Bertolini E, Olmos A, Vidal E, Martínez MC, Cambra M (2009) Direct sample preparation methods for the detection of *Plum pox virus* by real-time RT-PCR. *International Microbiology* **12**, 1–6.
- Chirkov S, Sheveleva A, Ivanov P, Zakubanskiy A (2018) Analysis of genetic diversity of Russian sour cherry *Plum pox virus* isolates provides evidence of a new strain. *Plant Disease* **102**, 569–575.
- Clemente-Moreno MJ, Hernández JA, Diaz-Vivancos P (2015) Sharka: how do plants respond to *Plum pox virus* infection? *Journal of Experimental Botany* **66**, 25–35.
- Crescenzi A, d'Aquino L, Comes S, Nuzzaci M, Piazzolla P, Boscía D, Hadidi A (1997) Characterization of the sweet cherry isolate of *Plum pox potyvirus*. *Plant Disease* **81**, 711–714.
- Dallot S, Bousalem M, Boeglin M, Renaud LY, Quiot JB (1997) Potential role of almond in sharka epidemics: susceptibility under controlled conditions to the main types of *Plum pox potyvirus* and survey for natural infections in France. *Bulletin OEPP/EPPO Bulletin* **27**, 539–546.
- Dallot S, Gottwald T, Labonne G, Quiot JB (2003) Spatial pattern analysis of sharka disease (*Plum pox virus* strain M) in peach orchards of southern France. *Phytopathology* **93**, 1543–1552.
- Damsteegt VD, Scorza R, Stone AL, Schneider WL, Webb K, Demuth M, Gildow FE (2007) *Prunus* host range of *Plum pox virus* (PPV) in the United States by aphid and graft inoculation. *Plant Disease* **91**, 18–23.
- Damsteegt VD, Waterworth HE, Mink GI, Howell WE, Levy L (1997) *Prunus tomentosa* as a diagnostic host for detection of *Plum pox virus* and other *Prunus* viruses. *Plant Disease* **81**, 329–332.
- Dosba F, Lansac M, Pêcheur G, Teyssier B, Piquemal JP, Michel M (1986) *Plum pox virus* detection by ELISA technique in peach and apricot infected trees at different growing stage. *Acta Horticulturae* **193**, 187–191.
- EPPO (1991/1992) Certification schemes. Virus-free or virus-tested fruit trees and rootstocks. *Bulletin OEPP/EPPO Bulletin* **21**, 267–278; **22**, 253–284.
- EPPO (2004) Diagnostic protocol for regulated pests. *Plum pox potyvirus*. *Bulletin OEPP/EPPO Bulletin* **34**, 247–256.
- EPPO (2016) Phytosanitary procedures. PM 3/76 (1) Trees of *Malus*, *Pyrus*, *Cydonia* and *Prunus* spp. – inspection of places of production. *Bulletin OEPP/EPPO Bulletin* **46**, 28–39.
- Fotiou IS, Pappi PG, Efthimiou KE, Katis NI, Maliogka VI (2019) Development of one-tube real-time RT-qPCR for the universal detection and quantification of *Plum pox virus* (PPV). *Journal of Virological Methods* **263**, 10–13.
- García JA, Glasa M, Cambra M, Candresse T (2014) *Plum pox virus* and sharka: A model potyvirus and a major disease. *Molecular Plant Pathology* **15**, 226–241.
- García JA, Cambra M (2007) *Plum pox virus* and sharka disease. *Plant Viruses* **1**, 69–79.
- Gentil P (2006) Detection of *Plum pox virus*: biological methods. *Bulletin OEPP/EPPO Bulletin* **36**, 251–253.
- Gildow F, Damsteegt V, Stone A, Schneider W, Luster D, Levy L (2004) *Plum pox* in North America: identification of aphid vectors and a potential role for fruit in virus spread. *Phytopathology* **94**, 868–874.
- Glasa M, Candresse T (2005) *Plum pox virus*. AAB Description of Plant Viruses. No. 410. <http://www.dpvweb.net/dpv/showdpv.php?dpvno=410>
- Glasa M, Hribovský I, Kúdela O (1999) Evidence for non-transmission of *Plum pox virus* by seed in infected plum

and myrobalan. *Biologia* **54**, 481-484.

Glasa M, Candresse T (2005) Partial sequence analysis of an atypical Turkish isolate provides further information on the evolutionary history of *Plum pox virus* (PPV). *Virus Research* **108**, 199-206.

Glasa M, Palkovics L, Komínek P, Labonne G, Pittnerova S, Kudela O, Candresse T, Šubr Z (2004) Geographically and temporally distant natural recombinant isolates of *Plum pox virus* (PPV) are genetically very similar and form a unique PPV subgroup. *Journal of General Virology* **85**, 2671–2681.

Glasa M, Predajna L, Šubr Z (2010) Competitiveness of different *Plum pox virus* isolates in experimental mixed infections reveals rather isolate- than strain specific behaviour. *Journal of Plant Pathology* **92**, 267-271.

Glasa M, Prikhodko Y, Predajna L, Nagyova A, Shneyder Y, Zhivaeva T, Subr Z, Cambra M, Candresse T (2013) Characterization of sour cherry isolates of *Plum pox virus* from the Volga basin in Russia reveals a new cherry strain of the virus. *Phytopathology* **103**, 972-979.

Gottwald TR, Avinent L, Llácer G, Hermoso de Mendoza A, Cambra M (1995) Analysis of the spatial spread of sharka (*Plum pox virus*) in apricot and peach orchards in eastern Spain. *Plant Disease* **79**, 266-278.

Hadersdorfer J, Neumüller M, Treutter D, Fischer TC (2011) Fast and reliable detection of *Plum pox virus* in woody host plants using the Blue LAMP protocol. *Annals of Applied Biology* **159**, 456-466.

Hadidi A, Levy L (1994) Accurate identification of *Plum pox potyvirus* and its differentiation from *Asian prunus latent potyvirus* in *Prunus* germplasm. *Bulletin OEPP/EPPO Bulletin* **24**, 633-643.

Hajizadeh M, Gibbs AJ, Amirnia F, Glasa M (2019) The global phylogeny of *Plum pox virus* is emerging. *Journal of General Virology* **100**, 1457-1468.

Hartmann W (1998) Hypersensitivity—a possibility for breeding sharka resistant plum hybrids. *Acta Horticulturae* **472**, 429–432.

Herrera G (2013) Investigations of the *Plum pox virus* in Chile in the past 20 years. *Chilean Journal of Agricultural Research* **73**, 60-65.

IPPC-FAO (2012) International standards for phytosanitary measures: diagnostic protocols: *Plum pox virus*. ISPM 27, Annex 2 (DP2).

James D, Thompson D (2006) Hosts and symptoms of *Plum pox virus*: ornamental and wild *Prunus* species. *Bulletin OEPP/EPPO Bulletin* **36**, 222-224.

James D, Varga A (2005) Nucleotide sequence analysis of *Plum pox virus* isolate W3174: evidence of a new strain. *Virus Research* **110**, 143-150.

Jridi C, Martin JF, Marie-Jeanne V, Labonne G, Blanc S (2006) Distinct viral populations differentiate and evolve independently in a single perennial host plant. *Journal of Virology* **80**, 2349-2357.

Kalashyan YA, Bilkey ND, Verderevskaya TD, Rubina EV (1994) *Plum pox potyvirus* on sour cherry in Moldova. *Bulletin OEPP/EPPO Bulletin* **24**, 645-649.

Kegler H, Fuchs E, Gruntzig M, Schwarz S (1998) Some results of 50 years of research on the resistance to *Plum pox virus*. *Acta Virologica* **42**, 200-215.

Kerlan C, Dunez J (1979) Différenciation biologique et sérologique des souches du virus de la sharka. *Annales de Phytopathologie* **11**, 241-250.

Kerlan C, Maison P, Lansac M, Dunez J (1980) Preliminary studies of the antagonism between strains of *Plum pox virus*. *Acta Phytopathologica Academiae Scientiarum Hungaricae* **15**, 57-68.

- Labonne G, Quiot JB (2001) Aphids can acquire *Plum pox virus* from infected fruits. *Acta Horticulturae* **550**, 79-84.
- Labonne G, Quiot JB (2006) The behaviour of alate aphids inside a *Prunus* orchard: an element to take into account for *Plum pox virus* spread? *Acta Horticulturae* **701**, 427-432.
- Labonne G, Yvon M, Quiot JB, Avinent L, Llacer G (1995) Aphids as potential vectors of *Plum pox virus*: comparison of methods of testing and epidemiological consequences. *Acta Horticulturae* **386**, 207-218.
- Levy L, Damsteegt V, Welliver R (2000) First report of *Plum pox virus* (sharka disease) in *Prunus persica* in the United States. *Plant Disease* **84**, 202.
- Llácer G (2006) Hosts and symptoms of *Plum pox virus*: herbaceous hosts. *Bulletin OEPP/EPPO Bulletin* **36**, 227-228.
- López MM, Bertolini E, Olmos A, Caruso P, Gorris MT, Llop P, Penyalver R, Cambra M (2003) Innovative tools for detection of plant pathogenic viruses and bacteria. *International Microbiology* **6**, 233-243.
- Maejima K, Hoshi H, Hashimoto M, Himeno M, Kawanishi T, Komatsu K, Yamaji Y, Hamamoto H, Namba S (2010) First report of *Plum pox virus* infecting Japanese apricot (*Prunus mume* Sieb. et Zucc.) in Japan. *Journal of General Plant Pathology* **76**, 229-231.
- Marais A, Faure C, Candresse T (2016) New insights into Asian *Prunus* viruses in the light of NGS-based full genome sequencing. *PLoS One* **11** (1): e0146420. doi:10.1371/journal.pone.0146420
- Martínez-Gómez P, Dicenta F, Audergon JM (2000) Behaviour of apricot (*Prunus armeniaca* L.) cultivars in the presence of sharka (*Plum pox potyvirus*): a review. *Agronomie* **20**, 407-422.
- Moreno A, Fereres A, Cambra M (2009) Quantitative estimation of *Plum pox virus* targets acquired and transmitted by a single *Myzus persicae*. *Archives of Virology* **154**, 1391-1399.
- Nemchinov L, Hadidi A, Maiss E, Cambra M, Candresse T, Damsteegt V (1996) Sour cherry strain of *Plum pox potyvirus* (PPV): molecular and serological evidence for a new subgroup of PPV strains. *Phytopathology* **86**, 1215-1221.
- Ng JC, Falk BW (2006) Virus-vector interactions mediating nonpersistent and semipersistent transmission of plant viruses. *Annual Reviews of Phytopathology* **44**, 183-212.
- Olmos A, Bertolini E, Cambra M (2002). Simultaneous and Co-operational amplification (Co-PCR) for detection of plant viruses. *Journal of Virological Methods* **106**, 51-59.
- Palmisano F, Boscia D, Minafra A, Myrta A, Candresse T (2012) An atypical Albanian isolate of *Plum pox virus* could be the progenitor of the Marcus strain. In: *22nd International Conference on Virus and Other Graft Transmissible Diseases of Fruit Crops*, June 3-8, Rome, Book of Abstracts, p. 33.
- Pasquini G, Barba M (2006) The question of seed transmissibility of *Plum pox virus*. *Bulletin OEPP/EPPO Bulletin* **36**, 287-292.
- Pleydell DRJ, Soubeyrand S, Dallot S, Labonne G, Chadœuf J, Jacquot E, Thébaud G (2018) Estimation of the dispersal distances of an aphid-borne virus in a patchy landscape. *PLoS Computational Biology* **14**(4): e1006085, doi: 10.1371/journal.pcbi.1006085
- Predajza L, Šubr Z, Candresse T, Glasa M (2012) Evaluation of the genetic diversity of *Plum pox virus* in a single plum tree. *Virus Research* **167**, 112-117.
- Rimbaud L, Dallot S, Delaunay A, Borron S, Soubeyrand S, Thébaud G, Jacquot E (2015b) Assessing the mismatch between incubation and latent periods for vector-borne diseases: The case of sharka. *Phytopathology* **105**, 1408-1416.

- Rimbaud L, Dallot S, Gottwald T, Decroocq V, Jacquot E, Soubeyrand S, Thébaud G (2015a) Sharka epidemiology and worldwide management strategies: learning lessons to optimize disease control in perennial plants. *Annual Reviews of Phytopathology* **53**, 357-378.
- Rodamilans B, Valli AA, Garcia JA (2019) Molecular Plant-*Plum pox virus* interactions. *Molecular Plant Microbe Interactions*, doi: 10.1094/MPMI-07-19-0189-FI (in press)
- Rodamilans B, Valli A, Mingot A, San León D, Baulcombe D, López-Moya JJ, García JA (2015) RNA polymerase slippage as a mechanism for the production of frameshift gene products in plant viruses of the Potyviridae family. *Journal of Virology* **89**, 6965-6967.
- Scorza R, Ravelonandro M, Callahan A, Zagrai I, Polak J, Malinowski T, Cambra M, Levy L, Damsteegt V, Krška B, Cordts J, Gonsalves D, Dardick C (2016) 'HoneySweet' (C5), the first genetically engineered plum pox (*Prunus domestica* L.) cultivar. *HortScience* **51**, 601–603.
- Sihelská N, Glasa M, Šubr Z (2017) Host preference of the major strains of *Plum pox virus* – opinions based on regional and world-wide sequence data. *Journal of Integrative Agriculture* **16**, 510-515.
- Sochor J, Babula P, Adam V, Krska B, Kizek R (2012) Sharka: the past, the present and the future. *Viruses* **4**, 2853-2901.
- Šubr Z, Glasa M (2008) *Plum pox virus* variability detected by the advanced analytical methods. *Acta Virologica* **52**, 75-90.
- Šubr Z, Pittnerova S, Glasa M (2004) A simplified RT-PCR-based detection of recombinant *Plum pox virus* isolates. *Acta Virologica* **48**, 173-176.
- Sutic D (1961) Assay of transmission of sharka virus disease by sap inoculation to herbaceous plants. *T. Planteavl.* **65**, 138-146.
- Thompson D, McCann M, McLeod M, Lye D, Green M, James D (2001) First report of *Plum pox potyvirus* in Canada. *Plant Disease* **85**, 97.
- Ulubas Serçe C, Candresse T, Svanella-Dumas L, Krizba, L, Gazel M, Çaglayan K (2009) Further characterization of a new recombinant group of *Plum pox virus* isolates, PPV-T, found in orchards in the Ankara province of Turkey. *Virus Research* **142**, 121-126.
- USDA (2019) USDA declares United States free from *Plum pox virus*. https://www.aphis.usda.gov/aphis/newsroom/news/sa_by_date/sa-2019/plum-pox-declaration
- Varga A, James D (2006) Use of reverse transcription loop-mediated isothermal amplification for the detection of *Plum pox virus*. *Journal of Virological Methods* **138**, 184-190.
- Varga A, James D (2005) Detection and differentiation of *Plum pox virus* using real-time multiplex PCR with SYBR Green and melting curve analysis: a rapid method for strain typing. *Journal of Virological Methods* **123**, 213-220.
- Vidal E, Zagrai L, Milusheva S, Bozhkova V, Tasheva-Terzieva E, Kamenova I, Zagrai I, Cambra M (2013) Horticultural mineral oil treatments in nurseries during aphid flights reduce *Plum pox virus* incidence under different ecological conditions. *Annals of Applied Biology* **162**, 299-308.
- Virsek Marn M, Mavric I, Urbancic-Zemljic M, Skerlavaj V (2004) Detection of *Plum pox potyvirus* in weeds. *Acta Horticulturae* **657**, 251-254.
- Wetzel T, Candresse T, Ravelonandro M, Delbos RP, Mazyad H, Aboul-Ata AE, Dunez J (1991) Nucleotide sequence of the 3' terminal region of the RNA of the El Amar strain of *Plum pox potyvirus*. *Journal of General Virology* **72**, 1741-1746.

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CABI/EPPO (1992/1997) *Quarantine Pests for Europe (1st and 2nd edition)*. CABI, Wallingford (GB).

EPPO (1983) Data sheets on quarantine organisms No. 96, *Plum pox virus*. *Bulletin OEPP/EPPO Bulletin* **13**(1), 1-7.